



Trophic Level Variations of Heavy Metals in Feathers of Birds from Awotan Landfill in Ibadan Nigeria

Ridwan Rotimi Abdulsalam^{1*}, Adeola Oni², and Aina Olubukola Adeogun²

¹Department of Biology, Federal University Otuoke, Otuoke, Bayelsa, Nigeria

²Department of Zoology, University of Ibadan, Ibadan, Oyo, Nigeria

Corresponding Author

Email: abdulsalamrr@fuio.edu.ng

Article Information	Abstract
<p>https://doi.org/10.69798/92699772</p> <p>ISSN (Online): 3066-3660</p> <p>Copyright ©: 2025 The Author(s). This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International (CC-BY-4.0) License, which permits the user to copy, distribute, and transmit the work provided that the original authors and source are credited.</p> <p>Published by: Koozakar LLC, Norcross GA 30071, United States. Note: The views expressed in this article are exclusively those of the authors and do not necessarily reflect the positions of their affiliated organizations, the publisher, the editors, or the reviewers. Any products discussed or claims made by their manufacturers are not guaranteed or endorsed by the publisher.</p> <p>Edited by: Oluseye Oludoye PhD</p>	<p>Global pollution from urbanization and technology exerts pressure on ecosystems through xenobiotics, especially heavy metals, which bioaccumulate and biomagnifies across food chains and trophic levels. In Nigeria, poorly managed landfills remain critical pollution hotspots, contributing to biodiversity loss and necessitating non-invasive ecotoxicological assessments. This study analyzed 13 metals (Fe, Mn, Zn, Cu, Co, Cr, Cd, Pb, Ni, Al, B, Se, Hg) in feathers of four bird species: <i>Hirundo aethiopica</i> and <i>Anthus leucophrys</i> (insectivores), <i>Streptopelia senegalensis</i> (granivore), and <i>Turdus pelios</i> (omnivore) using FAAS. Diversity indices and abundance were estimated via point counts, while ANOVA tested inter-trophic variation. Results showed elevated iron levels in insectivores (2.6985 ± 0.1975 ppm) compared to granivores (2.0100 ± 0.3172 ppm). Insectivores also accumulated higher levels of Cd, Co, Cr, Ni, Se, and Hg, whereas granivores consistently had lower concentrations. Significant differences were detected across trophic levels ($p < 0.05$). Findings indicate landfills serve as reservoirs of heavy metal pollution, with resident birds acting as effective sentinels of bioaccumulation, asymmetry, and ecosystem stress. Feather-based monitoring offers a reliable tool for tracking contaminant fate and predicting ecological risk.</p> <p>Keywords: Bioaccumulation and Biomagnification, Non-invasive Ecological Hazard Assessment, Toxic Heavy Metals, Avian Feathers, Ecological Declination and Stress</p>

INTRODUCTION

Environmental pollutants contaminate air, water, and soil, stressing ecosystems and biota. Human activities link closely to morbidity and mortality by impairing key biological functions (Usman et al., 2023). Poor solid waste management remains a major challenge (Alimba et al., 2006; Oshode et al., 2008) due to contaminants like heavy metals (Bakare et al., 2022). Urbanization and population growth increase waste generation, with landfills releasing leachates into groundwater, surface water, and gases into the atmosphere (Oshode et al., 2008). Unlike degradable pollutants, heavy metals are toxic, persistent, and prone to ecological accumulation with long half-lives. These include transition and post-transition metals (Nkwononwo et al., 2020), lanthanides, actinides (Adepoju-Bello et al., 2009), and metalloids (Igwe et al., 2005) characterized by high density (Tchounwou et al., 2012). Sources are both natural (ores, rocks) and anthropogenic: agriculture, metallurgy, energy production, atmospheric deposition, and waste discharge (Odika et al., 2020).

Some metals are trace nutrients, others strictly toxic (Ayeeni, 2014). Their bioavailability depends on pH, soil type, adsorption, and temperature, influencing toxicity at trace levels. Speciation affects solubility and transfer within soil, water, and sediments. Via bioaccumulation and biomagnification, metals disrupt ecosystems (Ali et al., 2019) and intensify at higher trophic levels. Non-biodegradability makes bioaccumulation problematic, as metals build up when uptake exceeds detoxification (Eagles-Smith et al., 2016), and birds are often used as bioaccumulative sentinels (Heys et al., 2016).

Monitoring heavy metals requires examining sources, persistence, and fate while soil, water, and air chemical monitoring reveals contamination (Pyagay et al., 2020), it often misses biological effects (Swaileh & Sansur, 2006). Biomonitoring using biological markers offers direct ecological insights (Rutkowska, 2018). Species and communities serve as pollution sentinels, reflecting ecosystem health, contamination pathways, and pollutant transfer through food webs (Gomez-Ramirez et al., 2021). Birds are particularly sensitive and widespread bioindicators of heavy metals. Due to ethical concerns over invasive tissue sampling (Acampora et al., 2017), researchers are shifting toward non-invasive methods (Garcia-Fernandez et al., 2013; Adeogun et al., 2022).

Feathers are prominent biomonitoring sentinels (Garcia-Fernandez et al., 2013). They reliably reflect internal metal concentrations, are easily collected and stored, and can be sampled across ages and sexes (Rutkowska et al., 2018). Feathers often contain higher metal loads

than other tissues (Malik & Zeb, 2009; Zamani-Ahmad et al., 2010) and are especially valuable for monitoring threatened species. Birds in anthropogenic landscapes are vulnerable, with feather contamination linked to developmental instability and dietary exposure (Malik & Zeb, 2009; Zamani-Ahmad Mahmoodi et al., 2010). Thus, feather-based assessments provide a reliable framework to evaluate toxic heavy metal (THM) accumulation and bioaccumulation patterns in avian trophic networks.

MATERIALS AND METHODS

Study Area

Ibadan, the capital of Oyo State, is one of Nigeria's largest cities, with approximately 3,080 sq. km and generating over 996,102 tons of solid waste annually (Amuda et al., 2014). The landfill lies 200–25 meters above sea level at latitude 07°27.59'N and longitude 03°50.93'E, along Apete-Awotan-Akufo Road, Apete, Ido Local Government Area, Ibadan. It receives municipal wastes from commercial, domestic, educational and industrial sources around the metropolis (Ogunseju et al., 2015).

Sampling

Bird surveys at the Awotan landfill were conducted between June and October 2021 following standard protocols (Edegbene, 2018). Sampling occurred weekly, with three visits in June and August, four in July, and one each in September and October. Both area counts and point counts were employed, the latter at three stations spaced 150 m apart: Point A (waste-dumping zone), Point B (vegetated area), and Point C (relatively an undisturbed site). Birds were observed for 5–20 minutes per station and identified to species level. Counts were done during peak activity periods: 06:30–11:00, 13:00–16:00, and 16:00–18:00 hrs. Observations took place while moving through the landfill and from elevated vantage points, with species confirmation aided by catalogues and identification manuals (Adeyanju et al., 2013). Complementary mist-netting followed procedures adapted from Adeyanju (2013).

Sampling points were randomly chosen considering avian habits and site characteristics. Three mist nets (20 × 10 m and 30 × 10 m, mesh sizes 25–45 mm) were positioned along likely flight paths or high-activity zones. Nets were installed at 06:30 hours and monitored every 10–20 minutes throughout the day, using improvised wooden poles for support.

Diversity Indices Analysis for Point Count Data

Margalef index (d)

It was used to measure the species richness of Awotan landfill using the formula below:

$$(d) = S-1 \div \ln N$$

Where;

d- Margalef index

S- total number of species

N- total number of individuals

Ln-Natural log

Shannon Weiner diversity indices (H)

The point count data of avian fauna encountered in Awotan was analyzed using the Shannon Weiner diversity indices state below:

$$H = -\sum PI \ln pi$$

Where;

PI- proportion of individual species

s- total number of species in the community

ln- natural logarithm

i- *i*th species

H- Shannon Weiner diversity

Pielou evenness index

It measures the evenness or equitability of the community and was determined using the formula below:

$$Evenness = H \div \log s$$

Where;

H- Shannon Weiner index

s- number of species/taxa

Log- Napierian log

Simpson's Diversity index (D)

It was used in comparing the diversity between avian fauna obtained from the point count data, taking into account species richness and evenness. It was calculated using the formula below:

$$D = \sum n(n-1) \div N-1$$

Where;

D-Dominance

n- Total number of individuals per species

N- Total number of organisms of all species counted in a point or station

Avian sampling

Birds were tagged using plastic avian leg rings to avoid recapturing and for future reference (Bergan et al., 2011). The captured birds (n=50) were weighed using electronic sf-400 and morphometric measurements (head length, beak/bill length, digit length, wing length, tail length and weight) were recorded. The total head length, beak/bill length, digit length, wing length, tail

length, and weight were taken from two individuals (n=2) of the Ethiopian swallow species (*Hirundo aethiopica*) to determine symmetry or asymmetry related to flight and morphometry. Two to four flight feathers from both wings were carefully removed from the right and left wings of the birds (Rutkowska et al., 2018) to avoid stressing or injuring the birds. Specific factors such as collection time, feather type, and feeding pattern were also noted (Garcia-Fernandez et al., 2013). The sampling process and analysis were non-invasive; birds were released after sampling, and collected feathers were stored in labeled Ziploc bags.

Sample Analysis: Feather Digestion and Metal Assay

Feathers were rinsed thrice with distilled water, oven-dried at 80°C (HVP-16426927), and homogenized using a ceramic mortar and pestle. Approximately 0.1 mg of powdered material was weighed (BA-T series analytical balance), digested in 2 mL of 70% HNO₃, and heated at 120°C for 24 hours. After cooling, samples were filtered (0.4 µm) and diluted to 25 mL with deionized water. Metal concentrations were measured using a Fast-Sequential Atomic Absorption Spectrophotometer (FAAS, Buck Scientific 210).

Statistical Analysis

One-way ANOVA was used to compare the differences in heavy metal concentrations in the feathers across various bird species and trophic levels at a significance level of $p < 0.05$.

RESULTS

A total of 702 individual birds, representing four families, were recorded at the Awotan landfill. These included two resident-migrant species (*Turdus pelios* and *Anthus leucophrys*) and two resident species. *Hirundo aethiopica* was the dominant species (Tables 1a, 1b). Mist-net captures yielded 50 birds, with *H. aethiopica* accounting for 74% (n=36), *Streptopelia senegalensis* 24% (n=12), and single captures of *Turdus pelios* and *Anthus leucophrys* (2% each) (Table 2). Species distribution by sampling point is shown in Table 3. Morphometric analyses included linear and mass measurements, with symmetry assessments conducted on *H. aethiopica* via bilateral wing, tail, and centrum comparisons (Tables 4a, 4b). Biodiversity metrics (Margalef, Shannon–Wiener, Simpson, and Pielou indices) indicated varying species abundance and diversity across sampling points (Tables 5a, 5b). Feather analyses detected 12 metals (Fe, Mn, Zn, Cu, Co, Cr, Cd, Ni, Al, B, Se, Hg), while lead (Pb) was undetected.

Concentrations varied among species and trophic levels (Tables 6, 7; Fig. 4a–c).

Table 1a: Area count checklist of avian species

Month	Common Name	Family	Scientific Name	Number Observed	% Occurrence
June	Ethiopian Swallow	Hirundinidae	<i>Hirundo aethiopica</i>	150	69.12
	Laughing Dove	Columbidae	<i>Streptopelia senegalensis</i>	65	29.95
	African Thrush	Turdidae	<i>Turdus pelios</i>	1	0.46
	Plain-Backed-Pipit	Motacillidae	<i>Anthus leucophrys</i>	1	0.46
			Total	217	100
July	Ethiopian Swallow	Hirundinidae	<i>Hirundo aethiopica</i>	120	58.25
	Laughing Dove	Columbidae	<i>Streptopelia senegalensis</i>	85	41.26
	African Thrush	Turdidae	<i>Turdus pelios</i>	1	0.49
			Total	206	100
August	Ethiopian Swallow	Hirundinidae	<i>Hirundo aethiopica</i>	135	62.5
	Laughing Dove	Columbidae	<i>Streptopelia senegalensis</i>	80	37.04
	African Thrush	Turdidae	<i>Turdus pelios</i>	1	0.46
			Total	216	100
September	Ethiopian Swallow	Hirundinidae	<i>Hirundo aethiopica</i>	8	50
	Laughing Dove	Columbidae	<i>Streptopelia senegalensis</i>	7	43.75
	African Thrush	Turdidae	<i>Turdus pelios</i>	1	6.25
			Total	16	100
October	Ethiopian Swallow	Hirundinidae	<i>Hirundo aethiopica</i>	35	74.47
	Laughing Dove	Columbidae	<i>Anthus leucophrys</i>	12	25.53
			Total	47	100

Table 1b: Summary table for the point count observation data

Common name	Family	Species Name	Number observed	% Occurrence
Laughing dove	Columbidae	<i>Streptopelia senegalensis</i>	249	35.47%
Ethiopian swallow	Hirundinidae	<i>Hirundo aethiopica</i>	448	63.82%
Plain-Backed-Pipit	Motacillidae	<i>Anthus leucophrys</i>	3	0.43%
African Thrush	Turdidae	<i>Turdus pelios</i>	2	0.29%
	Total		702	100%

Table 2: Mist net data of avian species encountered

Foraging behavior	Common name	Family	Species Name	Number observed	% Occurrence
Granivore	Laughing dove	Columbidae	Streptopelia senegalensis	12	24%
Insectivores	Ethiopian swallow	Hirundinidae	Hirundo aethiopica	36	72%
	Plain-Backed-Pipit	Motacillidae	Anthus leucophrys	1	2%
Omnivore	African Thrush	Turdidae	Turdus pelios	1	2%
		Total		50	100%

Species	N	Weight	Head Length	Beak	Full Tarsus	Digit	Wing
Ethiopian Swallow	36	12.610±2.678	3.831±0.322	0.600±0.209	0.273±0.431	1.247±0.258	9.600±0.498
Laughing dove	12	96.000±11.078	5.217±1.211	1.450±0.243	3.967±0.33	2.517±0.248	10.900±2.674
Plain-Backed-Pipit	1	32.000±0	4.600±0	1.200±0	5.000±0	2.000±0	9.100±0
African Thrush	1	68.000±0	5.600±0	1.700±0	5.100±0	2.700±0	10.500±0

Table 3: Avian species trapped in various sections

Points	Ethiopian Swallow	Laughing dove	African thrush	Plain-backed-Pipit	Total (point)
A	8	2	0	0	10
B	9	8	0	1	18
C	20	1	1	0	22
Total (species)	37	11	1	1	50

Table 4a: Morphometric measurements of Avian Species captured

Species	N	Weight	Head Length	Beak	Full Tarsus	Digit	Wing
Ethiopian Swallow	36	12.610±2.678	3.831±0.322	0.600±0.209	0.273±0.431	1.247±0.258	9.600±0.498
Laughing dove	12	96.000±11.078	5.217±1.211	1.450±0.243	3.967±0.33	2.517±0.248	10.900±2.674
Plain-Backed-Pipit	1	32.000±0	4.600±0	1.200±0	5.000±0	2.000±0	9.100±0
African Thrush	1	68.000±0	5.600±0	1.700±0	5.100±0	2.700±0	10.500±0

Values are given as mean ± standard deviation showing significance difference ($p < 0.05$)

Table 4b: Asymmetric measurements of the two Ethiopian Swallows

Morphometric Species	Right Wing Length	Left Wing Length	Symmetry	Left First Outer Tail	Right First Outer Tail	Symmetry	Centrum (Inner Tail Feather)
Ethiopian Swallow 1	9.9	9.9	Symmetric	5.8	5.9	Asymmetric	3.6
Ethiopian Swallow 2	10.4	10.5	Asymmetric	5.0	5.1	Asymmetric	3.9

Table 5a: Monthly biodiversity indices during the sampling period

Biodiversity indices	June	July	August
<i>Margalef Index (Species richness)</i>	0.668	0.379	0.758
<i>Shannon Weiner Index (Diversity)</i>	0.357	0.164	0.330
<i>Simpson index (Dominance)</i>	0.179	0.767	0.527
<i>Pielou Index (Evenness)</i>	0.357	0.136	0.288

Table 5b: Biodiversity indices for respective sampling points

Biodiversity indices	Point A	Point B	Point C
Margalef Index (Species richness)	0.435	0.692	0.647
Shannon Weiner Index (Diversity)	0.217	0.377	0.211
Simpson index (Dominance)	0.644	0.418	0.745
Pielou Index (Evenness)	0.217	0.300	0.157

Table 6: Mean concentration of heavy metals in analyzed feather samples

Metals ($\mu\text{g/ppm}$)	Species	Laughing Dove	Ethiopian Swallow	Plain-Backed-Pipit	African Thrush
	<i>Trophic level</i>	<i>Granivore</i>	<i>Insectivore</i>	<i>Insectivore</i>	<i>Omnivore</i>
	<i>Status</i>	<i>Resident</i>	<i>Resident</i>	<i>Resident-migrant</i>	<i>Resident-migrant</i>
	<i>N</i>	<i>3</i>	<i>3</i>	<i>2</i>	<i>2</i>
Manganese	0.633 \pm 0.0096		0.0643 \pm 0.0143	0.0900 \pm 0.0100	0.0585 \pm 0.0015
Iron	2.0100 \pm 0.3172		NA	2.6985 \pm 0.1975	NA
Zinc	0.1687 \pm 0.0110		0.1587 \pm 0.0195	0.1970 \pm 0.110	0.1350 \pm 0.0010
Cobalt	0.0007 \pm 0.0012		0.0003 \pm 0.0006	0.0005 \pm 0.0005	0.0015 \pm 0.0015
Chromium	0.0003 \pm 0.0006		NA	0.0005 \pm 0.0005	NA
Cadmium	0.0003 \pm 0.0006		NA	0.0005 \pm 0.0005	NA
Copper	0.0010 \pm 0.0010		NA	0.0010 \pm 0.0010	NA
Lead	ND		NA	ND	NA
Nickel	0.0003 \pm 0.0006		0.0007 \pm 0.0012	0.0005 \pm 0.0005	0.0010 \pm 0.0010
Aluminum	0.0357 \pm 0.0025		0.0287 \pm 0.0025	0.0240 \pm 0.0010	0.0495 \pm 0.0015
Boron	0.0137 \pm 0.0015		0.0180 \pm 0.0010	0.0185 \pm 0.0005	0.0245 \pm 0.0015
Selenium	0.0433 \pm 0.0015		0.0480 \pm 0.0017	0.0310 \pm 0.0020	0.0615 \pm 0.0035
Mercury	0.0213 \pm 0.0015		0.0270 \pm 0.0000	0.0120 \pm 0.0010	0.0330 \pm 0.0114

Values are given as mean \pm standard deviation showing significance difference ($p < 0.05$) of heavy metal concentrations in analyzed feathers

Note: NA - Not analyzed; ND -Not detected

Table 7: Mean concentration of heavy metals across trophic levels

Metals (μ/g-ppm)	Trophic level	Granivore	Insectivore	Omnivore
	Status	Resident	Resident-migrant	Resident-migrant
	N	3	5	2
Manganese		0.633±0.0096	0.0772±0.0122	0.0585±0.0015
Iron		2.0100±0.3172	2.6985±0.1975	NA
Zinc		0.1687±0.0110	0.1779±0.0153	0.1350±0.0010
Cobalt		0.0007±0.0012	0.0006±0.0009	0.0015±0.0015
Chromium		0.0003±0.0006	0.0005±0.0005	NA
Cadmium		0.0003±0.0006	0.0005±0.0005	NA
Copper		0.0010±0.0010	0.0010±0.0010	NA
Lead		ND	ND	ND
Nickel		0.0003±0.0006	0.0006±0.0015	0.0010±0.0010
Aluminum		0.0357±0.0025	0.0383±0.0023	0.0495±0.0015
Boron		0.0137±0.0015	0.0275±0.001	0.0245±0.0015
Selenium		0.0433±0.0015	0.0395±0.0027	0.0615±0.0035
Mercury		0.0213±0.0015	0.0125±0.0005	0.0330±0.0114

Values are given as mean ± standard deviation showing significance difference ($p < 0.05$) of heavy metal concentrations in birds feathers across trophic levels (granivore, insectivore and omnivore) from Awotan landfill

Note: NA - Not analyzed; ND - Not detected

DISCUSSION

Abundance and Diversity

The point count method evaluates avian relative abundance within habitats (Edegbeine, 2018). At the Awotan landfill, surveys revealed Ethiopian Swallows as the most prevalent species (63.82%, $n=448$), followed by Laughing Doves (35.47%, $n=249$), African Thrush (0.43%, $n=3$), and Plain-backed Pipit (0.28%, $n=2$). Monthly fluctuations in abundance were observed (Table 1a, 1b & 1c). Landfills often serve as resource-rich habitats due to continuous food disposal, which may explain the high bird presence (Osterback et al., 2015; Oka, 2016; Plaza & Lambertucci, 2017). Urban-adapted species such as Ethiopian Swallows display omnivorous or scavenging behaviors (Marasinghe et al., 2018). Species diversity is further influenced by anthropogenic activity, biotic interactions, and environmental heterogeneity (Protasov et al., 2009).

Diversity indices revealed spatial and temporal variation. Fifty individuals from four families were recorded across three sites: Point A (active dump), Point B (vegetated area without landfill activity), and Point C (inactive dump). Margalef's index was highest at Point B, moderate at Point C, and lowest at Point A (Table 5a, 5b). Shannon Wiener diversity peaked in June, declined in July, and increased again in August, with Point B consistently showing the greatest diversity. Simpson's dominance index

was highest at Point C, followed by Point A, while Point B exhibited the least dominance. July recorded the strongest dominance effect, suggesting reduced diversity consistent with pollution-driven resilience shifts where tolerant species outcompete sensitive taxa (Khan, 2016). Pielou's evenness indicated the most equitable species distribution in July, followed by August, with June being the least even. Spatially, Point B had the highest evenness, followed by Points A and C. Declines in evenness are linked to environmental stress, where pollutant-driven disturbances disrupt community balance (Faiz & Fakhar, 2016).

Overall, the Awotan landfill exhibited lower species richness and diversity relative to a stable local community. The observed variation in diversity and dominance likely reflects site-specific stresses and human impacts.

Heavy Metal Concentrations in Species

Heavy metal pollution in landfills has been widely reported, with evidence of bioaccumulation and biomagnification across trophic levels (Hammed et al., 2017; Marasinghe et al., 2018; Kinuthia et al., 2020). Previous research confirms that the Awotan landfill is contaminated, exposing resident biota, including birds, toxic metals (Olagunju et al., 2020; Oladejo et al., 2020; Adesogan & Omonigho, 2021). In this study, ten birds representing four species were analyzed (Table 6): Ethiopian Swallow ($n=3$), Laughing Dove ($n=3$), African

Thrush (n=2), and Plain-backed Pipit (n=2). Metal concentrations varied significantly among species and trophic levels ($p < 0.05$). Ethiopian Swallows exhibited the order $Zn > Mn > Se > Al > Hg > B > Ni > Co$, while Laughing Doves showed high Fe but negligible Pb (0.000 $\mu\text{g/g}$), following the pattern $Fe > Zn > Mn > Se > Al > Hg > B > Cu > Co > Cr/Cd/Ni > Pb$. African Thrushes recorded the highest Al, B, Se, Ni, and Hg levels, with the order $Zn > Se > Mn > Al > Hg > B > Co > Ni$. The Plain-backed Pipit demonstrated elevated Zn, Fe, and Mn but minimal Hg, Ni, Cd, and undetectable Pb, in the order $Fe > Zn > Mn > Se > Al > B > Hg > Cu > Cr/Cd/Co/Ni > Pb$. These interspecific variations reflect differences in diet and trophic ecology, with insectivores, granivores, and omnivores accumulating metals in distinct patterns (Jayakumar & Muralidharan, 2011). The findings underscore landfill-driven contamination and avifaunal vulnerability to heavy metal exposure across ecological niches.

Heavy Metal Concentrations across Avian Trophic Levels

Table 7 shows varying concentrations of heavy metals across trophic levels. High iron concentrations were detected, with insectivores (*Hirundo aethiopica*, *Anthus leucophrys*) showing the greatest levels, followed by granivores (*Streptopelia senegalensis*). Although within previously reported ranges (Einoder et al., 2018; Adesakin, 2021), levels exceeded safe limits (50–80 ppm), suggesting landfill inputs from metal and electronic wastes. Elevated iron may induce haemosiderosis and impair reproduction and moulting. Zinc, a common landfill contaminant (Alloway, 2005; Ngole & Ekosse, 2012), exceeded thresholds (Adesakin, 2021). Insectivores accumulated the most zinc, omnivores the least. Despite its metabolic role, excessive zinc is toxic. Mercury concentrations increased with trophic level, peaking in insectivores and lowest in granivores. Levels were below those reported by Keller et al. (2013), though some approached toxic thresholds (Evers et al., 2008). Mercury likely originates from electronics and industrial waste, bioaccumulating through insect or soil ingestion (Ab-Latif et al., 2015). Neurotoxic and hereditary risks remain high (Evers et al., 2008). Cadmium occurred at very low levels ($<2 \mu\text{g/g}$; Abdullah et al., 2015). Although insectivores showed higher accumulation, overall values suggest limited

contamination. Still, Cd exposure is linked to reproductive, renal, and developmental anomalies (Burger, 2008).

Cobalt and chromium were detected at low concentrations, though omnivores carried more cobalt. While cobalt is essential, excess amounts can impair thyroid and pulmonary health (Atashi et al., 2009). Chromium inputs, from industrial and waste sources (Jaishankar et al., 2014), were below thresholds but remain mutagenic (Patlolla et al., 2008). Nickel levels were low ($<5 \mu\text{g/g}$; Abdullah et al., 2015), with no trophic differences. Although widely used in alloys and electronics (Ngole & Ekosse, 2012), nickel exposure disrupts moulting, liver function, and immunity especially in avian species (Zivkov et al., 2017).

Manganese varied significantly across trophic levels, peaking in insectivores. Likely derived from fuel combustion and waste disposal, excess manganese can cause anemia, skeletal deformities, and behavioral disorders (Summers et al., 2011). Copper was below toxic thresholds (Jaynadeh et al., 2016), but insectivores accumulated the most. While essential, excess copper disrupts growth, endocrine, and reproductive functions (Stern, 2010).

Boron, mainly from detergents and agrochemicals, was highest in insectivores and lowest in omnivores. Aluminum was highest in omnivores, consistent with ingestion of contaminated soil or prey. Its oxidative stress effects are well documented (Slaninova et al., 2014). Selenium peaked in omnivores, with values ranging from below to above WHO thresholds (Adesakin, 2021). Likely sources include electronics, metallurgy, and landfill leachates (Mehdi et al., 2013). Chronic exposure can induce musculoskeletal defects and feather loss (Meschy, 2010). Lead was undetected, though its presence in landfill waste poses potential risks, including anemia, reproductive failure, and mortality in birds (Einoder et al., 2018).

Asymmetry and Asymmetric Morphometry of Birds

Ecosystem stability can be assessed through morphological indicators such as fluctuating asymmetry (FA), a subtle deviation from bilateral symmetry caused by developmental instability under stress (Lajus et al., 2015). Pollution, heavy metals, and persistent organic pollutants have been

strongly associated with increased FA and change of body physiology in wildlife, particularly birds (Jawad *et al.*, 2020). FA is widely used in biomonitoring because it reflects the combined influence of extrinsic stressors (contamination, habitat degradation) and intrinsic constraints (genetic load, inbreeding) on organismal development. In birds, wings and tail feathers are frequently used due to their aerodynamic and ecological significance. While natural asymmetry exists in primary feathers for flight efficiency, stress-induced asymmetry will reduce maneuverability, stability, and migration performance.

In this study, Ethiopian swallows (*Hirundo aethiopica*) from Awotan landfill were examined. Morphometric analysis showed asymmetry in wing length for the individual, but tail feathers showed asymmetry in both count and additional wing asymmetry in the second bird. Such deviations likely reflect environmental stresses from bioaccumulation, landfill contamination, consistent with earlier reports linking heavy metals especially mercury to avian feather asymmetry (Clarkson *et al.*, 2012). Asymmetry in feathers can alter flight energetics, prolong migration stops, and ultimately reduce survival in avian species. At the community level, such traits signal ecosystem degradation, making FA a sensitive biomonitoring tool and a potential early warning indicator for wildlife conservation (Cuervo & Retrepo, 2007).

CONCLUSION

Landfills act as critical drivers of ecological stress. Findings from Awotan landfill reveal substantial heavy metal pollution. Avian species exhibited bioaccumulation across trophic levels, underscoring both ecosystem vulnerability and the utility of birds as bioindicators. Feather based assays proved to be an efficient non-invasive biomonitoring tool, positing contamination patterns linked to trophism and waste-soil-food exposure. Notably metal loads in Ethiopian swallow species, points to associations with developmental instability and asymmetry, reinforcing the role of avian biomonitoring as an early warning system for environmental health and sustainability.

Data Availability

The data supporting this study are available from the corresponding author upon reasonable request.

Conflict of Interests

The authors declare no conflict of interest related to this study.

Funding Acknowledgement

No external funding was received for this study; it was supported by the authors' personal resources.

REFERENCES

- Abdullah, M., Fasola, M., Muhammad, A., Ahmad Malik, S., Bostan, N., Bokhari H., Aqeel Kamran, M., Nawaz Shafqat, M., A Alamdar, A., Khan, M., Ali, N., Ali Musstjab, S. & Shah Eqani, A. 2005. Avian feathers as a non-destructive bio-monitoring tool of trace metals signatures: A case study from severely contaminated areas. *Chemosphere*, 119, 553–561.
- Ab-Latif, W., Anjum, A. and Usmani, J.A. Lead toxicity: a review. *Interdisciplinary Toxicology*, 8 (2), 55-64.
- Acampora, H., White, P., Lyashevskaya, O., & O'Connor, I. 2017. Presence of Persistent organic pollutants in a breeding common tern (*Sterna hirundo*) population in Ireland. *Environmental Science and Pollution Research*, 25, 16933-16944.
- Ackerman, J.T., Eagles-Smith, C.A., Takekawa, J.Y., Bluso, J.D., & Adelsbach, T.L. 2008. Mercury concentrations in blood and feathers of prebreeding Forster's terns in relation to space use of San Francisco Bay, California, USA, habitats. *Environmental Toxicology and Chemistry*, 27 (4), 897–908.
- Adeogun, A.O., Chukwuka, A.V., Fadahunsi, A.A., Okali, K.D., Oluwakotanmi, P.G., Ibor, O.R., Emasoga, P. & Egware, T.U. 2022. Bird feathers as a non-invasive method for ecotoxicological monitoring: a rapid review. *The Zoologist*, 20, 26-40.
- Adepoju-Bello AA, Ojomolade OO, Ayoola GA, Coker AAB (2009). Quantitative analysis of some toxic metals in domestic water obtained from Lagos metropolis. *The Nigerian Journal of Pharmacy*, 42 (1): 57-60.
- Adesakin, T.A. 2021. Health hazards of toxic and essential heavy metals from thr poultry waste on human and aquatic organisms. *IntechOpen*
- Adesogan, S.O., and Omonigho, B.O. 2021. Assessment of leachate contamination potential of landfills in Ibadan, Nigeria. *African Journal of Environmental Science and Technology*, 15 (5), 179-187.
- Adeyanju, T. E., & Adeyanju, T. A. (2013). Avifauna of University of Ibadan Environs Ibadan, Nigeria *Proceedings of 3rd Annual Seminar of Nigerian Tropical Biological Association*. 27-34
- Ali, H., Khan, E., and Ilahi, I. 2019. Environmental chemistry and ecotoxicology of hazardous heavy metals: environmental persistence, toxicity, and bioaccumulation. *Journal of Chemistry*, 2019, (6730305).
- Alimba, C.G., Bakare, A.A., & Latunji, C.A. 2006 Municipal landfill leachates induced chromosome aberration in rat

- bone marrow cells. *African Journal of Biotechnology*, 5 (22), 2053-2057.
- Alloway, B.J. 2005. Copper-deficient soils in Europe. International Copper association. New York (2005)
- Amuda, O., Adebisi, S., Jimoda, L., Alade, A. 2014. Development Studies Challenges and Possible Panacea to the Municipal Solid Wastes Management in Nigeria. *Journal of Sustainable Development Studies*, 6.
- Atashi, H., Sahebi-Shahemabadi, F., Mansoorikiai, R. and Akbari, F. 2009. Cobalt in Zahedan drinking water. *Journal of Applied Sciences Research*, 5.
- Ayeni, O. 2014. Assessment of heavy metals in wastewater obtained from an industrial area in Ibadan, Nigeria. *RMZ – Materials and the Geoenvironment*, 61, 19–24.
- Bakare, A.A., Alimba, C.G. & Alabi, O.A. 2013. Genotoxicity and mutagenicity of solid waste leachates: A review, 12 (27), 4206-4220.
- Bergan, F., Endal, T., Lambert, F.M., Acosta Roa, A.M., Snartland, S., Steen, S., Steifetten, O., Sødning, M. & Aarvak, T., 2011. Guidelines for the identification of birds in research. Norwegian School of Veterinary Science. *Norecopa* (2011)
- Borrow, N. and Demey, R. 2014. *Birds of Western Africa*. 2nd Edition Volume 96 of Princeton Field Guides. Princeton University Press, 2014 ISBN-0691159203, 9780691159201.
- Clarkson, C.E., Erwin, R.M. & Riscassi, A. 2012. The use of novel biomarkers to determine dietary mercury accumulation in nestling waterbirds. *Environmental Toxicology and Chemistry*, 31, 1143–1148.
- Cuervo, A. & Restrepo, C. 2007. Assemblage and population-level consequences of forest fragmentation on bilateral asymmetry in tropical montane birds. *Biological Journal of the Linnean Society*. 92, 119–133.
- De L.S., Halitschek R., Hames, R.S., Kessler, A., De Voogd, T.J. & Dhondt, A.A. 2013. The Effect of Polychlorinated Biphenyls on the Song of Two Passerine Species. *PLoS ONE*, 8 (9).
- Eagles-Smith, C.A., Wiener, J.G., Eckley, C.S., Willacker, J.J., Evers, D.C., Marvin-Di Pasquale, M., Obrist, D., Fleck, J.A., Aiken, G.R., Lepak, J.M., Jackson, A.K., Webster, J.P., Stewart, A.R., Davis, J.A., Alpers, C.N. & Ackerman, T. 2016. Mercury in western North America: a synthesis of environmental contamination, fluxes, bioaccumulation, and risk to fish and wildlife. *Science and the Total Environment*, 568, 1213–1226.
- Edegbene, A.O. 2018. Invasive grass (*Typha domingensis*): A potential menace on the assemblage and abundance of migratory/water related birds in Hadeija-Nguru Wetlands, Yobe State, Nigeria. *Tropical and Freshwater Biology*, 27 (2), 13-20.
- Einoder, L.D., Southwell, D.M., Lahoz-Monfort, J.J., Gillespie, G.R., Fisher, A., Wintle, B.A. 2018. Occupancy and detectability modelling of vertebrates in northern Australia using multiple sampling methods. *PLoS ONE*, 13 (9).
- Evers, D.C., Savoy, L.J., DeSorbo, C.R., Yates, D.E., Hanson, W., Taylor, K.M., Siegel, L.S., Cooley Jr, J.H., Bank, M.S. & Major, A. et al. 2008. Adverse effects from environmental mercury loads on breeding common loons. *Ecotoxicology*, 17, 69-81.
- Faiz, Abu-UI and Fakhar-i-Abbas. 2016. Herpetofaunal Diversity of Tolipir National Park. *36th pakistan Congress of Zoology at: Lahore*.
- Garcia-Fernandez, A.J., Espin, S., Martinez-& Lopez, E. 2013. Feathers as a Biomonitoring Tool of Polyhalogenated Compounds: A Review. *Environmental Science & Technology*, 47, 3028-3043.
- Hammed, A.O., Lukuman, A., Gbola, K.A., & Mohammed, O.A. 2017. Heavy metal contents in soil and plants at dumpsites: A case study of Awotan and Ajakanga dumpsite Ibadan, Oyo State. Nigeria. *Journal of Environment and Earth Science*, 11-24.
- Heys, K., Shore, R., Pereira, M., Jones, K., & Martin, F. (2016). Risk assessment of environmental mixture effects. *Royal Society of Chemistry Advances*, 6.
- Jaspers, V.L.B., Voorspoels, S., Covaci, A., Lepoint, G. & Eens, M. 2007a. Evaluation of the usefulness of bird Feathers as a non-destructive biomonitoring tool for organic pollutants- a comparative and meta-analytical approach. *Environmental international*, APR, 33, 328-337.
- Janaydeh, M., Ismail, A., & Zulkifli, S.Z., Bejo, M.H., Abd. Aziz, N.A. & Taneenah, A. 2016. The use of feather as an indicator for heavy metal contamination in house crow (*Corvus splendens*) in the Klang area, Selangor, Malaysia. *Environmental Science and Pollution Research*, 23, 22059–22071.
- Jawad, L.A., Al-Janabi, M.I.G., & Rutkayová, J. (2020). Directional fluctuating asymmetry in certain morphological characters as a pollution indicator: Tigris catfish (*Silurus triostegus*) collected from the Euphrates, Tigris, and Shatt al-Arab Rivers in Iraq. *Fisheries & Aquatic Life*, 28 (1), 18-32.
- Jayakumar, R. and Muralidharan, S. 2011. Metal contamination in select species of fish and Nigiris district, Tamil Nadu, India. *Bulletin of Environmental Contamination and Toxicology*, 87 (2), 166-70.
- Khan, S.A. 2016. Tools for environmental Impact Assessment (EIA) using diversity indices. *Ving. Journal of Science*, 12 (1 & 2).
- Keller R.H., Xie L., Buchwalter D.B., Franzreb K.E. & Simons T.R. 2013. Mercury bioaccumulation in Southern Appalachian birds, assessed through feather concentrations. *Ecotoxicology*, 2013.
- Kinuthia, G.K., Ngure, V., Beti, D. 2020. Levels of heavy metals in wastewater and soil samples from open drainage channels in Nairobi, Kenya: community health implication. *Scientific Rep*, 10, 8434.
- Lajus, D., Knust, R., & Brix, O. (2003). Fluctuating asymmetry and other parameters of morphological variation of eelpout *Zoarces viviparus* (Zoaridae, Teleostei) from different parts of its distributional range. *Sarsia*, 88 (4), 247–260.
- Malik, R.N. & Zeb, N. 2009. Assessment of environmental contamination using feathers of *Bubulcus ibis* L., as a biomonitor of heavy metal pollution, Pakistan. *Ecotoxicology* 18 (5), 522–536.
- Marasinghe, S. Perera, P. & Dayawansa, N. 2018. Putrescible waste landfills as bird habitats in urban cities: A case from an urban landfill in the Colombo district of Sri Lanka. *Journal of Tropical Forestry and Environment*, 8.
- Martínez-López, E., Espín, S., Barbar, F., Lambertucci, S.A., Gómez-Ramírez, P., & García- Fernández, A. 2015.

- Contaminants in the southern tip of South America: Analysis of organochlorine compounds in feathers of avian scavengers from Argentinean Patagonia, *Ecotoxicology and Environmental Safety*, 115, 83–92.
- Mehdi, Y., Hornick, J.-L., Istasse, L. & Dufrasne, I. 2013. Selenium in the Environment, Metabolism and Involvement in Body Functions. *Molecules*, 18, 3292-3311.
- Meschy, F. *Nutrition minérale des ruminants*; Editions Quae: Versailles, France, **2010**; p. 208.
- Monclús, L., Lopez-Bejar, M., De la Puente, J., Covaci, A. & Jaspers, V.L.B., 2018. First evaluation of the use of down feathers for monitoring persistent organic pollutants and organophosphate ester flame retardants: A pilot study using nestlings of the endangered cinereous vulture (*Aegypius monachus*). *Environmental Pollution*, 238, 413-420.
- Ngole, V.M. & Ekosse, G.I.E. 2012. Copper, nickel and zinc contamination in soils within the precincts of mining and landfilling environments. *International Journal of Environmental Science and Technology*, 9, 485–494.
- Nkwunonwo, U.C., Odika, P.O., Onyia, N.I., 2020. A review of the health implications of heavy metals in food chains in Nigeria. *Scientific World Journal*, 30, 14.
- Ogunseiju, P., Ajayi, T.R., and Olarewaju, V.O. 2015. Geochemical assessment of the impact of an active open dumpsite on soil in Ibadan, Southwestern Nigeria. *Ife Journal of Science*, 17 (3). 12-19
- Oka, P.A. 2016. The Unnoticed Benefits of City Dumpsites. *Indian Journal of Applied Research*, 6, 113-119.
- Oladejo, O.P., Adagunodo, T.A., Sunmonu, L.A., Adabanija, M.A., Isibor, P.O., Enemuwe, C.A., 2020. Aeromagnetic mapping of fault architecture along Lagos-Ore axis, southwestern Nigeria. *Open Geosciences*, 10, 1515.
- Olagunju, T.E., Olagunju, A.O., Akawu, I.H., and Ugokwe, C.U., 2020. Quantification and Risk Assessment of Heavy Metals in Groundwater and Soil of Residential Areas around Awotan Landfill, Ibadan, Southwest Nigeria. *Journal of Toxicological Risk Assessment*, 6, 033.
- Omoyajowo, K. O., Njoku, K. L., Babalola, O. O., & Adenekan, O. A. (2017). Nutritional composition and heavy metal content of selected fruits in Nigeria. *Journal of Agriculture and Environment for International Development (JAEID)*, 111(1), 123–139.
- Oshode, O. A.1, Bakare, A. A., Adeogun, A. O., Efuntoye, M. O. & Sowunmi, A.A. 2008. Ecotoxicological Assessment Using *Clarias Gariepinus* and Microbial Characterization of Leachate from Municipal Solid Waste Landfill. *Int. Journal of Environmental Research*, 2 (4), 391-400.
- Osterback, A.-M.K., Frechette, D.M., Hayes, S.A., Shaffer, S.A. & Moore, J.W., 2015. Long-term shifts in anthropogenic subsidies to gulls and implications for an imperiled fish. *Biological Conservation*, 191, 606-613.
- Patlolla, A.K., Armstrong, N. and Tchounwou, P.B. 2008. Cytogenetic evaluation of potassium dichromate toxicity in bone marrow cells of Sprague-Dawley rats. *Metal Ions, Biology and Medicine*, 10, 353-358.
- Protasov, A., Barisov, S., Novoselova, T. & Syliaieva, A. 2019. The aquatic organism diversity, community structure, and the environmental conditions. *Diversity*, 11, 190.
- Pyagay, V.T., Sarsenova, Z.N., Duishekova, K.S., Duzbayev, N.T. & Albanbai, N. 2020. Analysis and processing of environmental monitoring system. *Procedia Computer Science*, 170, 26-33.
- Rutkowska, M., Płotka-Wasyłka, J., Lubinska-Szczygeł, M., Rozanska, A., Mozejko-Ciesielska, J. & Namiesnik, J. 2018. Birds' feathers- Suitable samples for determination of environmental pollutants. *Trends in Analytical Chemistry*, 109, 97-115.
- Slaninova, A., Machova, J. and Svobodova, Z. 2014. Fish kill caused by aluminium and iron contamination in a natural pond used for fish rearing: a case report. *Veterinarni Medicina*, 59 (11), 573-581.
- Stern, B.R. 2010. Essentiality and Toxicity in Copper Health Risk Assessment: Overview, Update and Regulatory Considerations, *Journal of Toxicology and Environmental Health*, 73 (2-3), 114-127.
- Summers, M., Summers, J., White, T. and Hannan, G. (2011). The effect of occupational exposure to manganese dust and fume on neuropsychological functioning in Australian smelter workers. *Journal of clinical and experimental neuropsychology*, 33.
- Swaleh, K. M. & Sansur, R. 2006. Monitoring urban heavy metal pollution using the House Sparrow (*Passer domesticus*). *Journal of Environmental Monitoring*, 8 (1), 209-213.
- Tchounwou, P.B., Yedjou, C.G., Patlolla, A.K., and Sutton, D.J. 2012. Heavy Metal Toxicity and the Environment. *Molecular, Clinical and Environmental Toxicology*, 101, 133-164.
- World Health Organization, 2011. Selenium in Drinking Water. *Background document for development*. WHO Guidelines for Drinking Water Quality.
- Zamani-Ahmadm Mahmoodi, R., Esmaili-Sari, A., Savabieasfahani, M., Ghasempouri, S.M. & Bahramifar, N., 2010. Mercury pollution in three species of waders from Shadegan wetlands at the head of the Persian Gulf. *Bull. Environmental Contamination and Toxicology*, 84, 326–330.
- Živkov Baloš, M., Ljubojević, D., & Jakšić, S. (2017). The role and importance of vanadium, chromium and nickel in poultry diet. *World's Poultry Science Journal*, 73 (1), 5–16.